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# Expanding the Phenotype Associated with NAA10-Related N-Terminal Acetylation Deficiency

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**ABSTRACT:** N-terminal acetylation is a common protein modification in eukaryotes associated with numerous cellular processes. Inherited mutations in NAA10, encoding the catalytic subunit of the major N-terminal acetylation complex NatA have been associated with diverse, syndromic X-linked recessive disorders, whereas de novo missense mutations have been reported in one male and one female individual with severe intellectual disability but otherwise unspecific phenotypes. Thus, the full genetic and clinical spectrum of NAA10 deficiency is yet to be delineated. We identified three different novel and one known missense mutation in NAA10, de novo in 11 females, and due to maternal germ line mosaicism in another girl and her more severely affected and deceased

brother. In vitro enzymatic assays for the novel, recurrent mutations p.(Arg83Cys) and p.(Phe128Leu) revealed reduced catalytic activity. X-inactivation was random in five females. The core phenotype of X-linked NAA10-related N-terminal-acetyltransferase deficiency in both males and females includes developmental delay, severe intellectual disability, postnatal growth failure with severe microcephaly, and skeletal or cardiac anomalies. Genotype–phenotype correlations within and between both genders are complex and may include various factors such as location and nature of mutations, enzymatic stability and activity, and X-inactivation in females.

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**KEY WORDS:** NAA10; X-linked; intellectual disability; N-terminal acetylation

Additional Supporting Information may be found in the online version of this article.

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## Introduction

N-terminal acetylation is a common protein modification in eukaryotes [Arnesen et al., 2009]. Recently, N-terminal acetylation was shown to act as a degradation signal to regulate protein complex stoichiometry, protein complex formation, subcellular targeting, and protein folding [Behnia et al., 2004; Setty et al., 2004; Forte et al., 2011; Scott et al., 2011; Shemorry et al., 2013; Holmes et al., 2014]. It is likely that the physiological role of N-terminal acetylation may vary, depending on the protein substrate. Six human

N-acetyltransferase enzymes (NATs) have been identified and designated NatA-NatF [Aksnes et al., 2015]. NatA has a major role and modifies approximately 40% of all human proteins [Arnesen et al., 2009]. It is composed of a catalytic subunit encoded by *NAA10* (ARD1; MIM #300013) (<http://www.ncbi.nlm.nih.gov/omim>) and an auxiliary subunit encoded by *NAA15* (NAT1/NATH; MIM #608000) [Mullen et al., 1989; Arnesen et al., 2005].

In recent years, mutations in the X-chromosomal *NAA10* gene have been implicated in various disease phenotypes. In 2011, Rope et al. identified the same missense mutation, p.(Ser37Pro) in two independent families with Ogden syndrome (MIM #300855), an X-linked recessive lethal disorder. Five affected males from one family and three affected males from the other family presented with a distinctive phenotype including postnatal growth failure, global severe developmental delay, an aged appearance with reduced subcutaneous fat, skin laxity, wrinkled foreheads, and facial dysmorphism such as prominent eyes, down-slanting palpebral fissures, thickened lids, large ears, flared nares with hypoplastic alae, short columella, protruding upper lip and microretrognathia. Furthermore, hypotonia progressing to hypertonia, cryptorchidism, hernias, large fontanels, and structural cardiac anomalies and/or arrhythmias were commonly noted. These individuals died between the ages of 5 and 16 months, mainly due to cardiac arrhythmias [Rope et al., 2011].

In 2014, *NAA10* was implicated in a second, distinct X-linked recessive disorder with syndromic microphthalmia (MCOPS1; MIM #309800). Esmailpour et al. identified the splice-site mutation c.471+2T>A in a family with Lenz microphthalmia syndrome. Several affected males presented with congenital bilateral anophthalmia, postnatal growth failure, hypotonia, skeletal anomalies such as scoliosis, pectus excavatum, finger, and toe syndactyly or clinodactyly, and with abnormal ears, fetal pads, and cardiac and renal anomalies. They had delayed motor development and mild to severe intellectual disability (ID) with remarkable intrafamilial variability. Furthermore, behavioral anomalies and seizures were reported. Three female carriers showed mild symptoms such as abnormally shaped ears and syndactyly of toes [Esmailpour et al., 2014].

Trio whole-exome sequencing within a larger study revealed the de novo missense mutation p.(Arg116Trp) in a boy with severe intellectual disability [Rauch et al., 2012]. Interestingly, shortly after another de novo missense mutation p.(Val107Phe) was identified in a female [Popp et al., 2015]. Both individuals with de novo mutations had severe cognitive impairment and showed variable other anomalies such as postnatal growth retardation, muscular hypotonia and skeletal, cardiac, and behavioral anomalies [Popp et al., 2015]. However, their phenotype was milder and less recognizable due to fewer malformations compared with the distinct syndromic entities of Ogden and syndromic microphthalmia syndromes [Popp et al., 2015]. Only very recently, another mutation in *NAA10*, p.(Tyr43Ser), was reported in two brothers with mild to moderate intellectual disability, scoliosis, and long QT and in their mildly affected mother [Casey et al., 2015].

Different levels of remaining enzymatic activity, but also the diversity of *NAA10* function, of which various aspects are potentially specifically targeted by different mutations, were discussed as underlying the various *NAA10* associated phenotypes [Esmailpour et al., 2014; Aksnes et al., 2015; Casey et al., 2015; Dorfel and Lyon, 2015; Popp et al., 2015]. However, given the small number of affected individuals and genetic variants identified as well as the lack of complete and comparative mechanistic investigations, no solid conclusion could yet be drawn.

We now report on a total of 12 affected females with four different de novo missense mutations in *NAA10* and one inherited mutation in a familial case due to germline mosaicism, thus further expanding

the mutational and clinical spectrum associated with *NAA10* related N-terminal-acetyltransferase deficiency.

## Material and Methods

### Individuals and Mutation Detection

All novel 11 affected females were seen and diagnosed in different centers worldwide. In all but individuals 4, 6, and 7, the mutation was detected by (trio) exome sequencing. Testing strategy in individual 5 has been recently published elsewhere [Thevenon et al., 2016]. In individuals 4, 6, and 7, the mutations were identified by targeted high-throughput sequencing of 275 genes implicated in intellectual disability (individual 6 and 7) as described elsewhere for a previous version of the sequencing approach [Redin et al., 2014] or by using the TruSight One sequencing panel by Illumina, containing approximately 4,800 disease associated genes (individual 4). In addition, we re-evaluated in detail a female with de novo mutation in *NAA10* who we recently reported (individual 1) [Popp et al., 2015]. Targeted mutation testing in the affected brother of individual 12 was done by Sanger sequencing. Analyses were performed either in a diagnostic or research setting, and informed consent was obtained from parents or guardians of all affected. If done in a research setting, the studies were approved by the ethic committees of the respective universities or centers.

Mutations and their de novo occurrence were confirmed by Sanger sequencing in the respective center. Gene version NM\_003491.3 was used as reference. Nucleotide numbering uses +1 as the A of the ATG translation initiation codon in the reference sequence, with the initiation codon as codon 1. The identified variants in *NAA10* were submitted to the LOVD gene variant database at <http://www.lovd.nl/NAA10> (individual IDs 60301—60312).

In silico prediction was performed with online programs SIFT [Kumar et al., 2009] (<http://sift.jcvi.org/>), PolyPhen-2 [Adzhubei et al., 2010] (<http://genetics.bwh.harvard.edu/pph2/>), and Mutation Taster [Schwarz et al., 2014] (<http://www.mutationtaster.org/>).

### X-inactivation

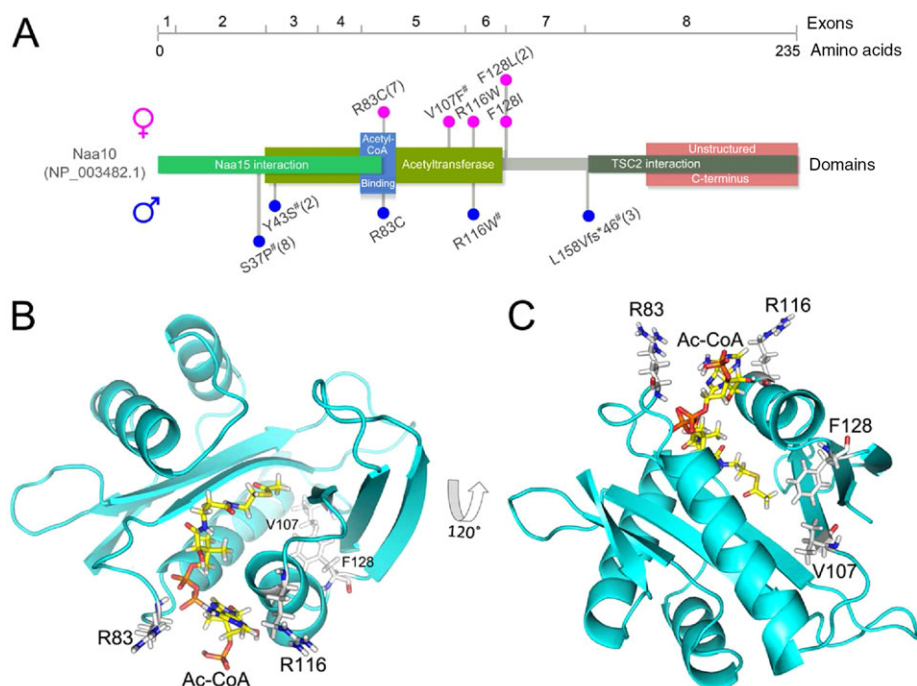
X-Inactivation testing at the androgen receptor locus was performed on lymphocytes of seven individuals according to standard procedures [Allen et al., 1992; Lau et al., 1997].

### Homology modelling

Structural consequences of the p.(Arg83Cys), the p.(Phe128Leu), and the p.(Phe128Ile) mutations were studied using a previously established homology model of the human *NAA10* enzyme based on the structural coordinates of *S. pombe* Naa10 [Myklebust et al., 2015] and visualized using MODELLER [Eswar et al., 2007].

### Plasmid construction and protein purification

Mutant His/MPB-*NAA10* and *NAA10*-V5 plasmids containing the p.(Arg83Cys), the p.(Phe128Leu), or p.(Phe128Ile) mutation were created by site-directed mutagenesis (QuikChange® Multi Site-Directed Mutagenesis Kit, Agilent Technologies, Santa Clara, CA) according to the manufacturer's protocol and as described previously [Popp et al., 2015] with the primers *NAA10*-R83C-Fwd: 5' GCGTTCCACCGGCGCCTCGGTCTGGCTCAGAACTGATGG and *NAA10*-R83C-Rev: 5' CCATCAGTTTCTGAGCCAGACCGAGGCGCCGGTGGGAACGC. Protein purification was performed



**Figure 1.** Mutations identified in *NAA10*. **A:** Schematic representation of the *NAA10* protein, encoding exons (numbered after NM\_003491.3), domains (based on NCBI reference NP\_003482.1, UniProt P41227 and the crystal structure of the NatA complex [Liszcak et al., 2013]), and localization of mutations in affected males (blue circles) and females (red circles). #, previously published mutations [Rauch et al., 2012; Rope et al., 2011; Esmailpour et al., 2014; Popp et al., 2015]. One letter codes were used due to space constraint. For the mutation c.471+2T>A described by Esmailpour et al. (2014) and the deduced protein level changes (p.(Leu158Valfs\*46), p.(Glu157\_Leu158ins9)), the presumably expressed truncated transcript 1 was used for labelling. **B:** *NAA10* homology model highlighting Arginine 83 and 116. These are localized in the Ac-CoA binding pocket. **C:** *NAA10* homology model showing the localization of Phe128, with its side chain pointing toward the hydrophobic core, similar to Val107. This amino acid was mutated in a previously published patient [Popp et al., 2015].

as described previously [Popp et al., 2015]. Buffers used for protein purification were: IMAC wash buffer (50 mM Tris-HCl [pH 7.4], 300 mM NaCl, 1 mM DTT, 25 mM Imidazole), IMAC elution buffer (50 mM Tris-HCl [pH 7.4], 300 mM NaCl, 1 mM DTT, 300 mM Imidazole), and size-exclusion chromatography buffer (50 mM HEPES [pH 7.5], 300 mM NaCl, 1 mM DTT). All enzymes that were purified and used for enzymatic acetylation assays were more than 95% pure.

### *In vitro* acetylation assays

A colorimetric acetylation assay [Thompson et al., 2004] and a quantitative HPLC assay [Evjenth et al., 2009] were used to measure the catalytic activity of the *NAA10* variants as described previously [Popp et al., 2015]. Purified enzyme was mixed with Acetyl-CoA (Ac-CoA), substrate peptides, and acetylation buffer (50 mM HEPES pH 7.5, 100 mM NaCl, and 2 mM EDTA), and reactions were carried out at 37°C. NAT activity was measured in the linear phase of the reaction. For  $K_m$  and  $k_{cat}$  determination, enzymes were mixed with 500  $\mu$ M oligopeptide and Ac-CoA ranging from 10  $\mu$ M to 1 mM. Due to the low catalytic activity of p.(Phe128Leu), kinetic constants were determined only for *NAA10* WT and *NAA10* p.(Arg83Cys).

### *Protein stability analysis for the p.(Arg83Cys) and the p.(Phe128Leu) mutations by cycloheximide chase assay*

Cycloheximide chase experiments were performed as described previously [Casey et al., 2015]. Briefly, HeLa cells were plated on six-

well plates and transfected with 2.6  $\mu$ g plasmid using X-tremeGENE nine DNA transfection reagent (Roche, Basel, Switzerland). Forty eight hours after transfection, 50  $\mu$ g/ml cycloheximide was added to the cells that subsequently were harvested at 0, 2, 4, and 6 hr post-treatment. V5-tagged *NAA10* variants were detected using a standard Western blotting procedure with anti-V5-tag antibody (R960-25; Invitrogen, ThermoFisher Scientific, Waltham, MA), anti- $\beta$ -Tubulin antibody (T5293; Sigma-Aldrich, St. Louis, MO), and anti-*NAA15* antibody (anti-NATH). Signals were detected by using ChemiDoc™ XRS+ system of Bio-RAD (Hercules, CA), and normalization and quantification of the signals was done with ImageLab™ system 5 of Bio-RAD. Each mutant was analyzed at least three times in independent experiments.

## Results

### Mutational Spectrum

Five different missense mutations in *NAA10* were identified in 12 female patients and one affected male (Fig. 1A; Supp. Table S1). The p.(Val107Phe) mutation has been reported previously [Popp et al., 2015] and is so far unique. Also, the p.(Arg116Trp) mutation has been reported previously, but in an affected boy [Popp et al., 2015]. Two mutations, p.(Phe128Ile) and p.(Phe128Leu), affect the same, highly conserved amino acid. The mutation p.(Phe128Ile) was identified in one female as recently reported [Thevenon et al., 2016], whereas p.(Phe128Leu) was identified in two unrelated female individuals. The recurrent mutation p.(Arg83Cys) was detected in seven females, one of them having an affected brother



who deceased at the age of 1 week. In this family, maternal germ line mosaicism was assumed because exome data did not reveal the mutation in the mother's blood sample and because of recurrence in a third pregnancy terminated after prenatal diagnosis. In all other 11 individuals, the mutation was shown to have occurred *de novo*.

None of the five missense mutations was found in the ExAC browser [Lek et al., 2015] (status November 2015) (<http://exac.broadinstitute.org/>). All affect highly conserved amino acids (Supp. Fig. S1 and [Popp et al., 2015]) and were classified as damaging or disease causing by Mutation Taster and SIFT and four of them as possibly or probably damaging by PolyPhen-2 (Supp. Table S2). X-inactivation was random in four tested individuals, mildly skewed (92%) in one and completely skewed (100%) in another.

## Clinical Phenotype

Clinical details of the affected individuals are displayed in Table 1. All 12 females are moderately to severely intellectually disabled. Age of walking ranges from 18 months to not being able to walk unassisted at age 8 years 6 months. Age of first words ranges from 15 months to no speech at age 8 years 6 months. Most of the individuals, though many still young, are nonverbal; one is reported to have a good vocabulary but not regularly using full sentences. Behavioral anomalies are described in seven girls and include stereotypic movements, self-hugging, attention deficit, aggressivity, and unmotivated laughter. Possible seizures or EEG anomalies were reported in only two of the individuals.

Birth measurements were in the normal or low normal range for most of the individuals. However, postnatal growth failure of length and/or head circumference occurred in most of the girls for whom this data were available. At age of last investigation (17 months–9 years), all but individuals 2 and 11 had head circumferences between –2.8 to –5.3 standard deviations (SD) below the mean. In relation, growth retardation regarding height was milder, and at age of last investigation length/height was in the normal or low normal range in seven individuals and between –2.5 and –3.75 SD in five. Feeding difficulties, either temporarily in infancy or continuously with the need of tube feeding, were reported in nine girls.

Muscular hypotonia was noted in almost all individuals, often more pronounced axially and with hypertonia of the extremities. Brain imaging, either by MRI or CNS ultrasound revealed frequent but rather unspecific anomalies. Structural cardiac anomalies such as pulmonary stenosis and atrium septum defects were observed in two females, cardiac conduction anomalies in five (twice long QT and three times incomplete right bundle branch block). Large fontanels were reported in four girls, and delayed bone age in one, but not tested in the others. Skeletal anomalies such as pectus excavatum or carinatum or vertebral anomalies were reported in five girls.

Various, but rather unspecific facial dysmorphism such as bitemporal narrowing, mildly up-slanting palpebral fissures, a prominent forehead, uplifted earlobes, hirsutism, or an up-turned nose were common. In several individuals, long eyelashes and arched, prominent eyebrows, or synophrys were noted (Fig. 2).

In one of the families, the p.(Arg83Cys) mutation also occurred in an affected brother due to maternal germ line mosaicism. This boy was born at gestational week 36 with normal birth length and weight. He had generalized hypotonia, lack of spontaneous respiration, and died at age 1 week. He had persistent pulmonary hypertension, supraventricular tachycardia, and mild ventricular hypertrophy. Ultrasound examination of the CNS showed ventriculomegaly, and small cortical kidney cysts were noted. Furthermore, he was

reported to have had large fontanels, bilateral hallux valgus, sandal gaps, a broad forehead, and micrognathia. A third, male pregnancy in this family was terminated after identifying the NAA10 mutation prenatally.

## Homology Modelling

The missense mutation p.(Arg83Cys) is located in the Ac-CoA binding site of NAA10, and the amino acid Arg83 might thus play an important role in the recruitment of Ac-CoA to the enzyme (Fig. 1B). The amino acids Arg83 and Arg116, the latter also mutated in a male individual (p.Arg116Trp) in an earlier study [Popp et al., 2015], are both positively charged and positioned on the rim of the Ac-CoA binding site. We thus suspect that the p.(Arg83Cys) mutation will have similar (or more adverse) effects as the p.(Arg116Trp) mutation previously identified. Phe128 on the other hand is not located in the active site of the enzyme (Fig. 1C). This residue is located in the loop region between  $\alpha 4$  and  $\beta 6$  of NAA10. The sidechain of Phe128 is pointing in toward a hydrophobic pocket between  $\beta 4$ ,  $\alpha 3$ ,  $\beta 5$ ,  $\alpha 4$ , and  $\beta 7$ . Previously, we described the p.(Val107Phe) variant whose sidechain is pointing toward the very same hydrophobic pocket and which resulted in more than 90% reduction of catalytic activity, most likely due to structural alterations or a reduced stability [Popp et al., 2015]. This indicates that the large bulky sidechain of Phe128 is necessary for this hydrophobic pocket and that the introduction of the much smaller Leucine or Isoleucine is causing a destabilization of the enzyme.

## *In vitro enzymatic assays and in cellula stability measurements*

In order to study the catalytic activity of NAA10 variants, plasmids coding for a His/MBP-NAA10 WT fusion protein, a His/MBP-NAA10 p.(Arg83Cys) fusion protein and a His/MBP-NAA10 p.(Phe128Leu) fusion protein were expressed in *E. coli* BL21 cells and purified. Interestingly, while NAA10 WT and NAA10 p.(Arg83Cys) eluted as monomers from the size-exclusion chromatography column, the NAA10 p.(Phe128Leu) eluted mainly in the void volume of the column (Supp. Fig. S2). This was also seen for another previously described NAA10 variant (p.(Tyr43Ser)) [Casey et al., 2015], and suggests that the protein has an altered protein structure or a reduced protein stability, causing it to aggregate in units larger than 200 kDa. For further enzymatic testing, only fractions of monomeric enzymes were used.

*In vitro* enzymatic assays revealed a drastic reduction of catalytic activity for NAA10 p.(Phe128Leu) (similarly to previously published NAA10 p.(Val107Phe) [Popp et al., 2015] and p.(Tyr43Ser) [Casey et al., 2015]), whereas the p.(Arg83Cys) mutation resulted in an approximately 60% reduction of catalytic activity (Fig. 3A). Together, these data support our findings from the homology model, suggesting that the p.(Phe128Leu) and p.(Phe128Ile) mutations interfere with the overall structure of NAA10 and possibly also destabilize the protein, whereas the p.(Arg83Cys) mutation interferes with Ac-CoA binding.

In order to further study whether the p.(Arg83Cys) mutation affects Ac-CoA, we determined steady-state kinetic constants (Supp. Fig. S3). The  $K_m$  for Ac-CoA is approximately threefold higher for the p.(Arg83Cys) variant compared with wild-type (WT) NAA10, indicating that the p.(Arg83Cys) variant has a reduced affinity toward Ac-CoA, and that this is the main cause of the reduced overall catalytic activity for this variant.

**Table 1. Clinical Details of Patients with De Novo Mutations in NAA10**

Individual	1 [Popp et al., 2015]	2	3	4	5 [Thevenon et al., 2016]	6	7	8	9	10	11	12*	13*
<i>NAA10</i> mutation	c.319G>T; p.(Val107Phe)	c.346C>T; p.(Arg116Trp)	c.384T>A; p.(Phe128Leu)	c.384T>A; p.(Phe128Leu)	c.382T>A; p.(Phe128Ile)	c.247C>T; p.(Arg83Cys)	c.247C>T; p.(Arg83Cys)	c.247C>T; p.(Arg83Cys)	c.247C>T; p.(Arg83Cys)	c.247C>T; p.(Arg83Cys)	c.247C>T; p.(Arg83Cys)	c.247C>T; p.(Arg83Cys)	c.247C>T; p.(Arg83Cys)
X-inact.	Random	Random	Random	NA	Random	Random	De novo	NA	NA	NA	100%	NA	NA
Inheritance	De novo	De novo	De novo	De novo	De novo	De novo	De novo	De novo	De novo	De novo	De novo	De novo	De novo
Mutation	Trio exome	Trio exome	Exome	Trio panel	Trio exome	Panel	Panel	Trio exome	Exome	Exome	Trio exome	Trio exome	Trio exome
detected by				(TS1)									
Gender	Female	Female	Female	Female	Female	Female	Female	Female	Female	Female	Female	Female	Female
Age last	8y 2mo	8y 1mo	17mo	8y 6mo	6y 5mo	4y 3mo	2y 2mo	3y 10mo	3y	10y 6mo	4y	6y 6mo	1wk/11wk
investigat.													
<b>Birth</b>													
Gestational week	Term	39+2	41	34	Term	Term	34 (twins)	38+1	37+1	37+1	40	41	36
Weight (g/SD)	Normal	3,350/0	3,020/-1.2	1,700/-1.3	2,670/-1.65	3,095/-0.67	2,210/0.2	2,540/-1.21	3,175/0.65	3,175/0.65	2,500/-2.31	3,470/-0.25	3,210/1.1
Length	Normal	49/-1	51/-0.13	NA	47/-1.51	47/-1.51	44/0	48/-0.3	52/1.72	52/1.72	49/-1.23	NA	NA
OFC (cm/SD)	NA	NA	32/-2.07	NA	32/-1.96	34/-0.51	30.5/0.1	32.4/-0.89	NA	NA	NA	NA	35.5/1.9
<b>Growth</b>													
Growth failure	Yes	No	No	Yes	Yes	Yes	Yes	Yes	Yes	Yes	No	Yes	NA
Height	108/-3.79	140/1.4	83.1/-0.92	115/-2.5	105/-2.65	97/-1.6	79.5/-2.56	88.3/-3.15	144/0.06	144/0.06	Normal	90/-1.62 (3y 2mo)	NA
Weight	17.5/-2.9	33.7/0.5	3/-0.39	17.2/-2.5	13.1/-4.12	12.6/-2.46	9.76/-2.26	11.2/-3.16	35/0.54	35/0.54	Normal	14/-0.35 (3y 2mo)	NA
(kg/SD)													
OFC (cm/SD)	48.5/-3.01	51/-0.3	41.7/-5.3	46/-4	46.2/-5.15	47/-3.59	44.7/-3.86	44.5/-4.85 (3y 5mo)	48 cm/-2.8 (6y)	48 cm/-2.8 (6y)	48.5/-1.27	45/-4.58 (3y 2mo)	NA
<b>CNS</b>													
Developm. delay/ID	Severe	Moderate (IQ ~ 50)	Severe	Profound	Yes	Yes	Severe	Severe	Severe	Moderate	Severe	Severe	NA
Age of walking	5y	18mo	Not yet	Not yet	Not yet	Not yet (3y 6mo)	Not yet	Not yet	Not yet	3y	24mo	Not yet	NA
First words/current speech	Not yet/signs and sounds	15mo	Not yet/consonant babbles	Not yet	Not yet	2y (2 words)	Not yet	Not yet	Not yet	3y / good, but not always full sentences	Not yet	Not yet, sounds	NA
Seizures (age)	No	No	Infantile spasms (4mo)	5y	No	No	No	No	"Spells" vs. seizures	No	No	No	NA
EEG anomalies		NA	Myoclonic, generalized	Bifrontal Axial		NA	No	No	No	No	No	No	Yes
Hypotonia/hypertonia	Axial hypo/periph hyper	Hypo	Severe hypo	hypo/periph hyper	Axial hypo/periph hyper	Hypo	Hypo (axial > periph)	Hypo	Hypo/hyper	Hypo	No	Periph hyper	Hypo
Brain imaging anomalies	US normal	MRI normal (6y)	MRI (9mo): parenchymal atrophy, thin CC	Normal (1y)	Supraventricular cyst without hydrocephaly	Hippocampic dysgenesis	MRI (20mo): periventric. white matter loss	IVH occipital horn, PVL, HIE	MRI normal (12mo, 3y)	MRI normal (12mo, 3y)			
Borderline ventricu-lomegaly	MRI: thin CC	Dilated lat. ventricles, thin CC	US (pre- and perinatal): ventricu-lomegaly										

(Continued)

**Table 1. Continued**

Individual	1 [Popp et al., 2015]	2	3	4	5 [Thevenon et al., 2016]	6	7	8	9	10	11	12*	13*
Behavioral anomalies	Active, short attention span, friendly	ADHD	No	Stereotypies, hand washing, uncertain eye contact	Incomplete right bundle branch block	Unmotivated laughter	Aggressivity	No	Happy disposition	Very active, happy, problems in new settings	Hyperactivity, little eye contact, aggressivity	Restlessness, attention deficit, happy disposition	NA
<b>Visceral</b>													
Cardiac anomalies	PAS, ASD, long QT (max QT time 484 ms)	Incomplete right bundle branch block	Mild left ventr. diastolic dysfunction, ECG normal	No	Incomplete right bundle branch block	No	No	Mild PAS, PFO vs. ASD	No	Long QT	Incomplete right bundle branch block	No	SVT, pulm. hypert., mild ventric. hypertrophy
Eye anomalies	Astigmatism, strabism, mild optic atrophy	No	Cortical vision impairment	Cortical visual impairment, amblyopia	Hyperopia, astigmatism, exotropia	No	No	Myopia, astigmatism	Alternating esotropia, cortical vision impairment	Astigmatism, hyperopia	Hyperopia	Megalopapillae, myopia	NA
Urogenital/kidney anomalies	No	NA	NA	NA	No	NA	No	NA	No	Prenatal renal fluid	No	Small cysts unilateral	Diffuse small cortical kidney cysts
<b>Skeletal</b>													
Large fontanels	Yes	No	No	NA	NA	No	NA	Yes	Yes	Yes	No	No	Yes
Delayed bone age	Yes	NA	NA	No	NA	NA	NA	NA	NA	NA	NA	NA	NA
Hand anomalies	No	No	No	No	No	No	No	No	Small hands	No	Clinodactyly V	Small hands, tapering fingers	NA
Feet anomalies	Broad great toes	No	No	No	No	No	No	No	Small	No	No	Small feet and toes	Hallux varus, sandal gap
Thoracic anomalies	Mild pectus carinatum	No	No	No	No	No	No	Supernumerary vertebra, dysmorphic L1 vertebra	Yes	Mild pectus excavatum	Pectus excavatum	Pectus excavatum	NA
<b>Other</b>													
Feeding difficulties	Yes, infancy	No	Yes, g-tube	Yes, tube in infancy, choking	Yes	Yes	No	Yes, infancy	Yes	Mild, in infancy	Yes, infancy	NA	NA
Other anomalies or difficulties	Recurrent infections, high pain threshold	DCD		Pyramidal signs, dysonic postures		Narrow palate	VP shunt, high palate	Resolved cutis marmorata	Sleeping problems	Cutis marmorata			

\*siblings.

MGM, maternal germline mosaicism; anom., anomalies; TS1, Trisight One sequencing panel (Illumina); †, deceased; mut., mutation; mo, months; y, years; wk(s), week(s); NA, not available/applicable; OFC, occipitofrontal head circumference; CNS, central nervous system; ID, intellectual disability; US, ultrasound; CC, corpus callosum; US, ultrasound; hyop, hypotonia; hyper, hypertonia; periph, peripheral; ADHD, attention deficit hyperactivity disorder; PAS, pulmonary artery stenosis; ASD, atrial septum defect; SVT, supraventricular tachycardia; US, ultrasound; DCD, (developmental coordination disorder); HIE, hypoxic-ischemic encephalopathy; PVL, periventricular leukomalacia; VP, ventriculo-peritoneal.



**Figure 2.** Facial phenotypes of females with NAA10-related N-terminal-acetyltransferase deficiency. Note only minor dysmorphic aspects and a rather unspecific facial gestalt in most of the girls. Some have prominent or arched eyebrows.

We also assessed the stability of the NAA10 variants by over-expression in HeLa cells followed by cycloheximide-chase experiments. The NAA10 p.(Phe128Leu) variant was less stable compared with NAA10 WT (Fig. 3B–E). Similar findings were observed for NAA10 Phe128Ile (data not shown). In contrast, protein stability does not seem to be affected by the p.(Arg83Cys) mutation.

## Discussion

### Delineation of the Phenotype in Females with NAA10-Related N-Terminal-Acetyltransferase Deficiency

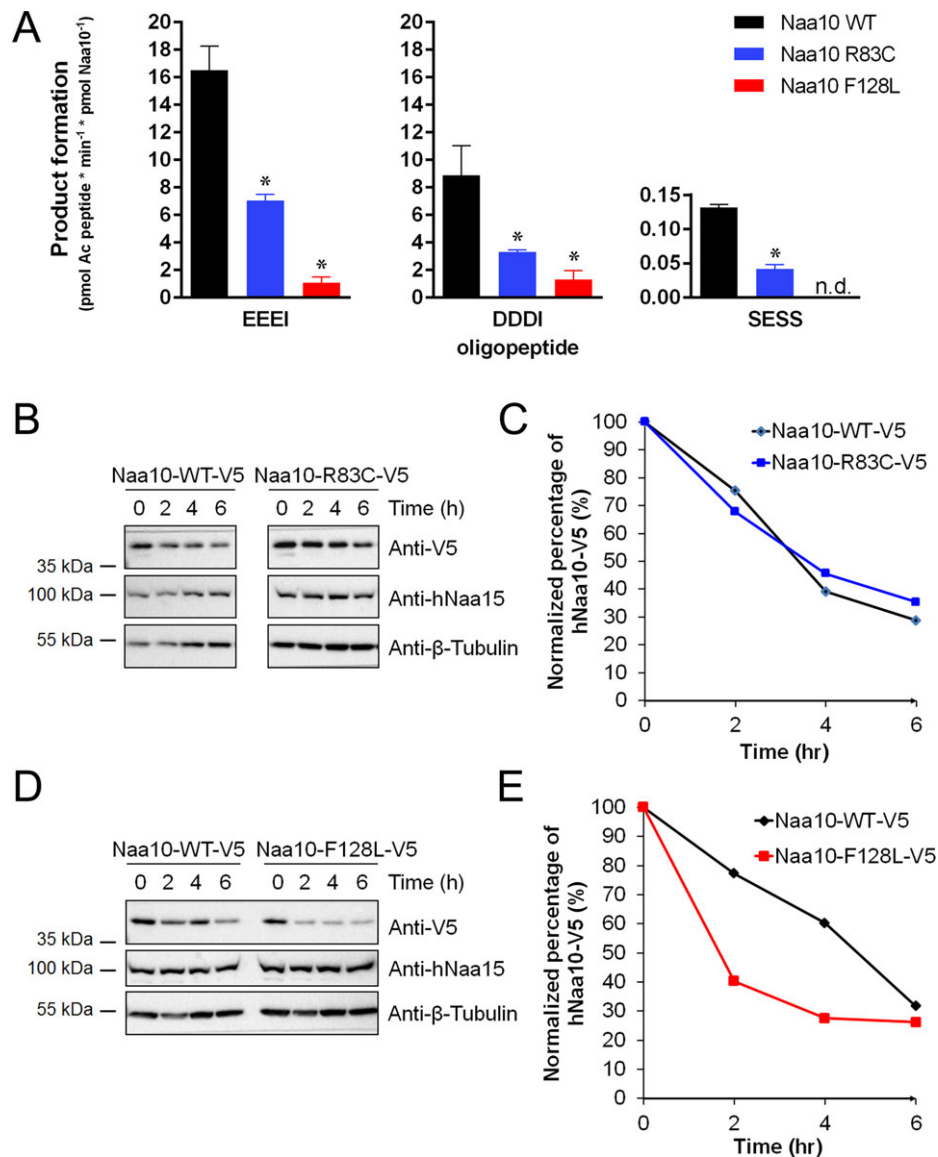
We describe 12 female individuals, 11 of whom carry de novo missense mutations in *NAA10*, a gene previously implicated in different X-linked recessive disorders. The herewith reported females all have moderate to severe ID and variable other anomalies, partly overlapping with those in the affected males but with a generally milder and less characteristic phenotype.

Several common clinical aspects could be noted among the affected females. All had moderate to severe developmental delay and ID with no or very limited speech and often with limited mobility. While birth measurements were often in the normal or low normal range, postnatal growth failure commonly occurred, also resulting in severe microcephaly. Skeletal, brain, and organ anomalies were frequent, but without any apparent specific pattern. Minor, but often rather unspecific facial dysmorphism were observed in most of the patients. In several individuals, arched eyebrows or synophrys were noted, which, if present, in combination with short

stature and intellectual disability might bear some resemblance to Cornelia-de-Lange syndrome (CDLS1; MIM #122470, CDLS2; MIM #300590, CDLS3; MIM #610759, CDLS4; MIM #614701, CDLS5; MIM #300882). As the p.(Arg83Cys) mutation was recurrently identified in seven girls, we wondered whether it might be accompanied by specific phenotypic aspects. However, we could not deduce any obvious differences in clinical appearance between individuals with this mutation or one of the other missense mutations.

Importantly, cardiac conduction anomalies were observed in five of the females. This might be a milder manifestation of the cardiac arrhythmias that were frequent and often lethal in many of the boys with Ogden syndrome [Rope et al., 2011]. Also, very recently, long-QT was observed in two male individuals and their mother with NAA10-related N-terminal acetylation deficiency [Casey et al., 2015]. As the observation of conduction defects has important consequences, such anomalies should be specifically tested in newly identified individuals. Furthermore, specific recommendations for surveillance, medication, or avoidance of risk factors should be given to families with affected children. As long as little information on the course of NAA10-related N-terminal-acetyltransferase deficiency in females is available, general recommendations for cardiac conduction anomalies such as long-QT syndrome might be applied. One should also consider electrocardiographic examination in otherwise asymptomatic female carriers in families with X-linked recessive NAA10-related N-terminal-acetyltransferase deficiency, as conduction anomalies might occur as a minimal manifestation of the disease in them, similar to cardiomyopathies in female carriers of Duchenne muscular dystrophy [Hoogerwaard et al., 1999].





**Figure 3.** Functional consequences of *NAA10* mutations. **A:** Purified wild-type (WT) or mutant *NAA10* were each mixed with acetylation buffer, 500  $\mu$ M peptide and 500  $\mu$ M Ac-CoA, and incubated at 37°C. Reactions were stopped in the linear phase of the reaction. The p.(Arg83Cys) mutation resulted in an approximately 60% reduction in catalytic activity of *NAA10* for all tested oligopeptides, whereas the p.(Phe128Leu) mutation lead to a very low catalytic activity (more than 90% reduction). **B–E:** *NAA10* stability in vivo, tested pairwise (mutant and control) by cyclohexamide chase experiments and monitored at different time points (between 0 and 6 hr post-treatment) and by Western blotting. Figures show representative results from at least three independent experiments. **B:** Western blot for *NAA10*-V5 WT or *NAA10*-V5 carrying the mutation p.(Arg83C). Bands were quantified, and the relative amount of *NAA10* present at each time point is showed in **(C)**. **D:** Western blot for *NAA10*-V5 WT or *NAA10*-V5 carrying the mutation p.(Phe128Leu). Bands were quantified, and the relative amount of *NAA10* present at each time point is showed in **(E)**.

## Genotype–Phenotype Correlations

Developmental delay and intellectual disability, postnatal growth failure, and cardiac and skeletal anomalies are overlapping clinical aspects between male individuals with X-linked recessive Ogden syndrome and syndromic microphthalmia and with males or females with severe ID due to de novo mutations in *NAA10*. These features therefore probably represent general clinical consequences of *NAA10*-related N-terminal acetylation deficiency. We therefore suggest using this or the shorter term “*NAA10* deficiency” to describe the associated phenotypes instead of splitting them into different syndromic entities. The more specific aspects such as aged appearance or wrinkled skin in Ogden syndrome [Rope

et al., 2011], anophthalmia, seizures, and syndactyly in syndromic microphthalmia [Esmailpour et al., 2014] or a rather mild phenotype in two brothers with mild to moderate ID, scoliosis, and long-QT [Casey et al., 2015] might be related to specific effects of the respective, causative mutations.

Different levels of remaining enzymatic activity and specifically affected properties of *NAA10* function were reported for different *NAA10* mutations [Esmailpour et al., 2014; Aksnes et al., 2015; Casey et al., 2015; Dorfel and Lyon, 2015; Popp et al., 2015]. In the family with syndromic microphthalmia due to a splice-site mutation (c.471+2T>A), absent expression of WT *NAA10* in fibroblasts derived from affected individuals and only minimal expression of mutant *NAA10* was shown [Esmailpour et al., 2014].

For the p.(Ser37Pro) mutation from the Ogden syndrome families, disruption of the catalytic activity was confirmed [Rope et al., 2011]. Furthermore, the integrity of the NatA complex was disturbed and N-terminal acetylome analyses of B-cells, and fibroblasts from affected individuals revealed a reduction of the NatA-mediated N-terminal acetylation [Van Damme et al., 2014; Myklebust et al., 2015]. In contrast, the de novo p.(Arg116Trp) mutation resulted in a high remaining catalytic activity, therefore being in accordance with the relatively mild phenotype in the affected boy [Popp et al., 2015], whereas the de novo p.(Val107Phe) mutation, identified in a girl, had a more severe effect with almost abolished catalytic activity, thus explaining the pronounced manifestation of symptoms in a female [Popp et al., 2015].

However, the hypothesis of phenotypic severity correlating with levels of catalytic activity is challenged by a report on two affected brothers with a rather mild phenotype but carrying a mutation with a severe impact on catalytic activity [Casey et al., 2015].

Our functional studies, comparing all de novo missense mutations identified in females, show that the p.(Phe128Leu) and the p.(Phe128Ile) mutations cause a reduced stability of NAA10 and loss of NAA10 NAT-activity. Based on the close proximity of Phe128 and Val107 in the NAA10 structure, our data suggest that individuals with either mutation might be affected through the same molecular mechanisms. The p.(Arg83Cys) mutation also has a clearly reduced catalytic activity, due to a lower affinity for AC-CoA, but does not seem to affect NAA10 structurally. The homogenous phenotype in all girls with either the p.(Val107Phe), p.(Phe128Leu), p.(Phe128Ile), or p.(Arg83Cys) mutation suggests that rather the loss of the NAT activity, which is shared by all missense mutations, and not the impaired structural stability, found for all but the p.(Arg83Cys) mutation, is underlying their similar phenotype. The milder catalytic impairment by the p.(Arg83Cys) mutation of 60% compared to a >90% abolishment of catalytic activity by p.(Phe128Leu) or the previously published p.(Val107Phe) variant is not reflected in an obviously milder clinical phenotype in the respective individuals, suggesting a critical threshold of enzyme activity necessary for normal function. This critical threshold might, however, be met by individual 2. She showed a milder phenotype with an IQ of 50 and no postnatal growth deficiency. This girl carries the mutation p.(Arg116Trp), that has previously been shown to result in a rather mild reduction of NAA10 catalytic activity [Popp et al., 2015].

According to the observation of the p.(Arg116Trp) mutation causing a more severe phenotype in a male [Popp et al., 2015] compared to the herewith reported female, also the other missense mutations might have a significantly more pronounced effect in males, possibly leading to lethality. This would be in accordance with the observation in family 12, in which the affected boy deceased 1 week after birth. Nevertheless, the contrast between the severe ID and microcephaly phenotypes in the herewith reported females with mainly de novo mutations in NAA10 and the mostly asymptomatic carrier females in families with Ogden and Lenz microphthalmia syndromes is still striking. A possible cause might be the X-inactivation pattern. Its possible impact on the phenotype is reflected in Individual 11. This girl has an 100% skewed X-inactivation in lymphocytes and is the only individual carrying the p.(Arg83Cys) mutation but not showing microcephaly or postnatal growth retardation. In contrast, X-inactivation was random in lymphocytes of four of the other five tested females in our study and only mildly skewed in the fifth. All carrier females in the original Ogden syndrome family were reported to have skewed X-inactivation, which would be in accordance with them not showing any or only mild symptoms [Myklebust et al., 2015]. However, an explanation why skewing of X-inactivation oc-

curred in the females with inherited mutations and only twice in the females with de novo mutations and still causing a severe phenotype, is still lacking. As NAA10-related N-terminal acetyltransferase deficiency seems to be caused by different mutations and to be accompanied by various phenotypes, there might be tissue specificity or a correlation between specific mutations and skewing or nonskewing of X-inactivation.

### *De novo mutations in X-linked recessive genes in females*

The relatively high number of 12 individuals indicates that missense mutations affecting the catalytic activity of X-chromosomal encoded NAA10 might be a relatively frequent cause of severe ID in females. Similar observations of *de novo* mutations in X-linked genes causing severe ID phenotypes in females while female carriers of mutations in families with affected males were normal, were recently made for other genes such as *DDX3X* (MIM# 300160) [Snijders Blok et al., 2015] or *PHF6* (MIM# 300414) [Zweier et al., 2013]. We therefore hypothesize that *de novo* mutations in supposedly X-linked recessive genes might be more common than previously expected.

To conclude, our findings show that different missense mutations in NAA10 in females result in severe ID and postnatal growth failure with severe microcephaly and thus expand the clinical and mutational spectrum associated with X-linked NAA10 related N-terminal-acetyltransferase deficiency. Particularly the observation of cardiac conduction anomalies in both males and females has relevant consequences for the clinical management of known and newly identified individuals.

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### **References**

- Adzhubei IA, Schmidt S, Peshkin L, Ramensky VE, Gerasimova A, Bork P, Kondrashov AS, Sunyaev SR. 2010. A method and server for predicting damaging missense mutations. *Nat Methods* 7:248–249.
- Aksnes H, Hole K, Arnesen T. 2015. Molecular, cellular, and physiological significance of N-terminal acetylation. *Int Rev Cell Mol Biol* 316:267–305.
- Allen RC, Zoghbi HY, Moseley AB, Rosenblatt HM, Belmont JW. 1992. Methylation of HpaII and HhaI sites near the polymorphic CAG repeat in the human androgen-receptor gene correlates with X chromosome inactivation. *Am J Hum Genet* 51:1229–1239.
- Arnesen T, Anderson D, Baldersheim C, Lanotte M, Varhaug JE, Lillehaug JR. 2005. Identification and characterization of the human ARD1-NATH protein acetyltransferase complex. *Biochem J* 386:433–443.
- Arnesen T, Van Damme P, Polevoda B, Helsens K, Evjenth R, Colaert N, Varhaug JE, Vandekerckhove J, Lillehaug JR, Sherman F, Gevaert K. 2009. Proteomics analyses reveal the evolutionary conservation and divergence of N-terminal acetyltransferases from yeast and humans. *Proc Natl Acad Sci U S A* 106:8157–8162.
- Behnia R, Panic B, Whyte JR, Munro S. 2004. Targeting of the Arf-like GTPase Arl3p to the Golgi requires N-terminal acetylation and the membrane protein Sys1p. *Nat Cell Biol* 6:405–413.
- Casey JP, Stove SI, McGorrian C, Galvin J, Blenski M, Dunne A, Ennis S, Brett F, King MD, Arnesen T, Lynch SA. 2015. NAA10 mutation causing a novel intellectual disability syndrome with Long QT due to N-terminal acetyltransferase impairment. *Sci Rep* 5:16022.

- Dorfel MJ, Lyon GJ. 2015. The biological functions of Naa10—from amino-terminal acetylation to human disease. *Gene* 567:103–131.
- Esmailpour T, Riazifar H, Liu L, Donkervoort S, Huang VH, Madaan S, Shoucri BM, Busch A, Wu J, Towbin A, Chadwick RB, Sequeira A, et al. 2014. A splice donor mutation in NAA10 results in the dysregulation of the retinoic acid signalling pathway and causes Lenz microphthalmia syndrome. *J Med Genet* 51:185–196.
- Eswar N, Webb B, Marti-Renom MA, Madhusudhan MS, Eramian D, Shen MY, Pieper U, Sali A. 2007. Comparative protein structure modeling using MODELLER. *Curr Protoc Protein Sci Chapter 2:Unit 2:9*.
- Evjenth R, Hole K, Ziegler M, Lillehaug JR. 2009. Application of reverse-phase HPLC to quantify oligopeptide acetylation eliminates interference from unspecific acetyl CoA hydrolysis. *BMC Proc* 3 Suppl 6:55.
- Forte GM, Pool MR, Stirling CJ. 2011. N-terminal acetylation inhibits protein targeting to the endoplasmic reticulum. *PLoS Biol* 9:e1001073.
- Holmes WM, Mannakee BK, Gutenkunst RN, Serio TR. 2014. Loss of amino-terminal acetylation suppresses a prion phenotype by modulating global protein folding. *Nat Commun* 5:4383.
- Hoogerwaard EM, Bakker E, Ippel PF, Oosterwijk JC, Majoor-Krakauer DF, Leschot NJ, Van Essen AJ, Brunner HG, van der Wouwe PA, Wilde AA, de Visser M. 1999. Signs and symptoms of Duchenne muscular dystrophy and Becker muscular dystrophy among carriers in The Netherlands: a cohort study. *Lancet* 353:2116–2119.
- Kumar P, Henikoff S, Ng PC. 2009. Predicting the effects of coding non-synonymous variants on protein function using the SIFT algorithm. *Nat Protoc* 4:1073–1081.
- Lau AW, Brown CJ, Penaherrera M, Langlois S, Kalousek DK, Robinson WP. 1997. Skewed X-chromosome inactivation is common in fetuses or newborns associated with confined placental mosaicism. *Am J Hum Genet* 61:1353–1361.
- Lek M, Karczewski K, Minikel E, Samocha K, Banks E, Fennell T, O'Donnell-Luria A, Ware J, Hill A, Cummings B, Tukiainen T, Birnbaum D, et al. 2015. Analysis of protein-coding genetic variation in 60,706 humans. *bioRxiv*. <http://dx.doi.org/10.1101/030338>
- Liszcak G, Goldberg JM, Foyn H, Petersson EJ, Arnesen T, Marmorstein R. 2013. Molecular basis for N-terminal acetylation by the heterodimeric NatA complex. *Nat Struct Mol Biol* 20:1098–1105.
- Mullen JR, Kayne PS, Moerschell RP, Tsunasawa S, Gribskov M, Colavito-Shepanski M, Grunstein M, Sherman F, Sternglanz R. 1989. Identification and characterization of genes and mutants for an N-terminal acetyltransferase from yeast. *EMBO J* 8:2067–2075.
- Myklebust LM, Van Damme P, Stove SI, Dorfel MJ, Abboud A, Kalvik TV, Grauffel C, Jonckheere V, Wu Y, Swensen J, Kaasa H, Liszcak G, et al. 2015. Biochemical and cellular analysis of Ogden syndrome reveals downstream Nt-acetylation defects. *Hum Mol Genet* 24:1956–1976.
- Popp B, Stove SI, Ende S, Myklebust LM, Hoyer J, Sticht H, Azzarello-Burri S, Rauch A, Arnesen T, Reis A. 2015. De novo missense mutations in the NAA10 gene cause severe non-syndromic developmental delay in males and females. *Eur J Hum Genet* 23:602–609.
- Rauch A, Wieczorek D, Graf E, Wieland T, Ende S, Schwarzmayr T, Albrecht B, Bartholdi D, Beygo J, Di Donato N, Dufke A, Cremer K, et al. 2012. Range of genetic mutations associated with severe non-syndromic sporadic intellectual disability: an exome sequencing study. *Lancet* 380:1674–1682.
- Redin C, Gerard B, Lauer J, Herenger Y, Muller J, Quartier A, Masurel-Paulet A, Willems M, Lesca G, El-Chehadeh S, Le Gras S, Vicaire S, et al. 2014. Efficient strategy for the molecular diagnosis of intellectual disability using targeted high-throughput sequencing. *J Med Genet* 51:724–736.
- Rope AF, Wang K, Evjenth R, Xing J, Johnston JJ, Swensen JJ, Johnson WE, Moore B, Huff CD, Bird LM, Carey JC, Opitz JM, et al. 2011. Using VAAST to identify an X-linked disorder resulting in lethality in male infants due to N-terminal acetyltransferase deficiency. *Am J Hum Genet* 89:28–43.
- Schwarz JM, Cooper DN, Schuelke M, Seelow D. 2014. MutationTaster2: mutation prediction for the deep-sequencing age. *Nat Methods* 11:361–362.
- Scott DC, Monda JK, Bennett EJ, Harper JW, Schulman BA. 2011. N-terminal acetylation acts as an avidity enhancer within an interconnected multiprotein complex. *Science* 334:674–678.
- Setty SR, Strohlic TI, Tong AH, Boone C, Burd CG. 2004. Golgi targeting of ARF-like GTPase Arl3p requires its Nalpa-acetylation and the integral membrane protein Sys1p. *Nat Cell Biol* 6:414–419.
- Shemorry A, Hwang CS, Varshavsky A. 2013. Control of protein quality and stoichiometries by N-terminal acetylation and the N-end rule pathway. *Mol Cell* 50:540–551.
- Snijders Blok L, Madsen E, Juusola J, Gilissen C, Baralle D, Reijnders MR, Venselaar H, Helmsmoortel C, Cho MT, Hoischen A, Vissers LE, Koemans TS, et al. 2015. Mutations in DDX3X are a common cause of unexplained intellectual disability with gender-specific effects on Wnt signaling. *Am J Hum Genet* 97:343–352.
- Thevenon J, Duffourd Y, Masurel-Paulet A, Lefebvre M, Feillet F, El Chehadeh-Djebbar S, St-Onge J, Steinmetz A, Huet F, Chouchane M, Darmency-Stamboul V, Callier P, et al. 2016. Diagnostic odyssey in severe neurodevelopmental disorders: towards clinical whole-exome sequencing as a first-line diagnostic test. *Clin Genet* 10.1111/cge.12732
- Thompson PR, Wang D, Wang L, Fulco M, Pediconi N, Zhang D, An W, Ge Q, Roeder RG, Wong J, Levrero M, Sartorelli V, et al. 2004. Regulation of the p300 HAT domain via a novel activation loop. *Nat Struct Mol Biol* 11:308–315.
- Van Damme P, Stove SI, Glomnes N, Gevaert K, Arnesen T. 2014. A *Saccharomyces cerevisiae* model reveals in vivo functional impairment of the Ogden syndrome N-terminal acetyltransferase NAA10 Ser37Pro mutant. *Mol Cell Proteomics* 13:2031–2041.
- Zweier C, Kraus C, Brueton L, Cole T, Degenhardt F, Engels H, Gillessen-Kaesbach G, Graul-Neumann L, Horn D, Hoyer J, Just W, Rauch A, et al. 2013. A new face of Borjeson-Forssman-Lehmann syndrome? De novo mutations in PHF6 in seven females with a distinct phenotype. *J Med Genet* 50:838–847.